

Innate and Adaptive Immunity in Fish: A Review

Enas Sheet Mustafa and Shahbaa Khalil AL-Tae

Department of Pathology and Poultry Diseases, Collage of Veterinary Medicine, University of Mosul, Iraq

Corresponding author; shahbaa_khal@uomosul.edu.iq, ORCID:0000-0001-7798-7091

Doi: <https://doi.org/10.37940/AJVS.2020.13.2.1>

Received: 8/4/2020 Accepted:4/7/2020

This article is licensed under a CC BY (Creative Commons Attribution 4.0)

<http://creativecommons.org/licenses/by/4.0/>.

Abstract

The fish inhabitant in an aquatic environment so it persists exposure to pathogen and stressor factors so they have a developing immune system similar to that in high vertebrate with some differences points. Head Kidney considered the main and primary hematopoietic organs while spleen, thymus and gut-associated lymphoid tissue represented secondary hematopoietic organs. The immune system divided to the innate immune system doesn't have a memory of previous responses, non-specific cellular as natural killer cells, dendritic cells, neutrophils, basophils and eosinophils with non-specific component these involve complement, lysosomes and acute-phase protein, if the pathogen persists the specific, memory adaptive immunity would be stimulation which composed of two subunits humeral and natural antibodies act for invading extracellular pathogen and the second subunits which are cytokines and T-lymphocyte act for kill intracellular bacterial, parasitic and viral infection. Both subunits (innate and adaptive) of the immune system act each together in the hosts to prevent microorganism's infection and reducer the environmental stressors and give fish immune status. It is concluded from this article review that fish, like mammalians, have an advanced immune system that plays a role in fish resistance to pathological factors and maintaining fish health.

Keywords: Fish, Immunity, Innate, Adaptive.

المناعة الفطرية غير المتخصصة والتكيفية المتخصصة في الاسماك

الخلاصة

تعيش الأسماك في البيئة المائية ولذا فانها تتعرض الى المسببات المرضية وعوامل الاجهاد وعليه تمتلك جهاز مناعي متطور مماثل لذلك الموجود في الفقاريات العالية مع بعض نقاط الاختلاف. تعد الكلية الراسية من مكونات الأعضاء اللمفاوية الرئيسية والأولية بينما يمثل النسيج الليمفاوي المرتبط بالطحال والتوتة والأمعاء الأعضاء المكونة للدم الثانوية. يقسم الجهاز المناعي الى المناعة الفطرية غير المتخصصة والذي لا يتميز بذاكرة للاستجابات السابقة وخلايا غير محددة كالخلايا القاتلة الطبيعية والخلايا المتشجرة والعدلات والقاعدية والحمضة ومكونات غير متخصصة والتي تشمل المكمل والحالات وطور البروتين الحاد إذا استمر العامل الممرض ، فإن المناعة التكيفية المتخصصة للذاكرة ستكون عبارة عن تحفيز يتكون من وحدتين فرعيتين من الأجسام المضادة العضدية والطبيعية تعمل على غزو مسببات الأمراض خارج الخلية والوحدات الفرعية الثانية وهي السيتوكينات واللمفاويات التائية تعمل لقتل العدوى البكتيرية والطفيلية والفيروسية داخل الخلايا. تعمل كل من الوحدتين الفرعيتين (الفطرية والتكيفية) من الجهاز المناعي معاً في المضيف لمنع عدوى الكائنات الحية الدقيقة وتقليل الضغوطات البيئية وإعطاء حالة مناعة الأسماك. يستنتج من مراجعة المقالة هذه بان الأسماك كالبائت لها جهاز مناعي متقدم يلعب دور مهم في مقاومة الأسماك للعوامل الامراضية والمحافظة على صحة الأسماك.

Introduction

Both animals (vertebrates and invertebrate) with plants have immunosystem (1) which play important function in protect themselves against pathogens(2).The immune system in fish is physiologically and functionally similar to that higher vertebrate which are mainly produce immune cells by cytokines and associated receptors is the main function against pathogen and protecting organisms against diseases (3).

There was some variable features, in fish there is no bone marrow which represented the primary immune organs in other vertebrates, head kidney(HK) in fish represented the organ for hematopoietic cells , myeloid , lymphoid immune cells and produced antibodies (Ab), (4) thymus represented center for T cell maturation (5), also fish lack lymph nodes, blood infiltration is done by Melan macrophage centers which is characterized by assembly macrophages combined to ellipsoid capillaries (5).

Ontogeny of Immune System

The lymphoid tissue is the main unit of immune system in fish because fish lack lymph nodes and bone marrow, although there is 24.0000 fish species was morphological differences but the lymph tissue mainly consisting from reticular cells providing the innate and adaptive immunity as dendritic cells, mononuclear cells, granulocyte, mast cells, thrombocyte and natural killer cells (6).

The first hematopoietic organ in teleost fish was known as intermediated cells Mas (ICM), in some fish species as Kili (*Pseudoepiplatys annulatus*) and Rain Bow Trout hematopoietic persisted for short period and then shift to ICM (7).The lymphoid organs are anterior kidney, thymus, spleen and gut associated lymphoid organs (GALT) were completed before

embryonic hatching. (8) refer to variable sequence development of lymphoid organs in marine fish which as kidney ,spleen and thymus with variable function related to stage as spleen in the larval stage has erythropoietic function than hematopoietic. (9) Determined the observative time for each cell, myeloid cell persist around the yolk sac from 24 hours post fertilization(hpf) but not are seen in ICM, neutrophils and myeloblast observation and migrate through blood vessels walls during 34 and 48hpf while granulocytes are presented in blood supply in the connective tissue surround the yolk sac at 34 hpf and at 48 hpf observative near renal tubules.

Immunoglobulins (IgM) and B lymphocyte varies among fish species, they are late appearance in marine fish compare to freshwater fish. IgM play roles in phagocytosis, activation complement and it represented nutritional yolk protein (10).

In teleost fish as in higher vertebrate the maternal Abs act to protect eggs against vertical transfer pathogen (11).

Lymphoid Organs

Spleen

The mammal's spleen consist from 2-part white pulp composed mainly from hematopoietic for defense mechanism and red pulp that causes phagocytosis for abnormal and old cells. These features may variable from fish spleen although it persisting in some spp. The fish spleen composed from melanomacrophage (MMC), lymphoid tissue and surrounding with clustered ellipsoid (12), macrophage phagocytize antigen that will culminate in immune memory and then represented for T lymphocyte (13), in zebra fish, spleen describes as small organ contain a large number of enitroblast at 30day post fertilization at three month the lymphoblast antigen , these developmental pattern has been

detected in Catfish, Grouper and Atlantic salmon(14; 15)

Thymus

It has two lobes mainly consist of thin sheet of oval lymphoid tissue located behind operculum in the dorsolateral position of gill lined with mucosa of pharyngeal epithelium (16), origin of the thymus occur at 24hpf, aggregation of macrophage which are promote encapsulated proliferation T lymphocytes and the capsule surrounding lymphoid tissue are the two main component of fish thymus these structure is variable according to fish in contrast to higher vertebrate it is difficult to recognized medulla from cortex(17).

Thymus responsible for the production and maturation of T lymphocyte as well as the eosinophilic granulocytes and myeloid cells are detected in thymus with Hassal,s corpuscles (7).

Kidney

The kidney in teleost fish are recognized as two lobes anterior and posterior kidney has play roles in blood filtration,urine formation and secretion hormone as renin and angiotensin while anterior kidney equivalent of the bone marrow in vertebrate it is important for production defense cells, reorganization and maturation of B lymphocytes, monocyte and granulocytes(18)

Fish Immune System

As in human and other vertebrates the immune system divided in to two subunit, innate (non-specific) immunity composed of three component (physical barrier, humeral and cellular factors), innate immunity recognised the pathogenic organisms(lipopolysaccharide LPS)in gram negative bacteria, lipoteichoic acid (LTA) in

gram positive bacteria, beta -glucan and chitin in fungi and hemagglutinin in viruses by pathogenic associated molecular patterns (PAMPs) (19; 20) after that the phagocytosis process occur, if pathogen persist for long time the innate immunity replace by second subunit adaptive immunity which is more specific and Igs, T cells receptors with major histocompatibility are the main component of it (21; 22).

Innate Immunity

Physical Barrier

In aquatic environment fish is usually direct contact with many types of microorganisms, so the main and first line defense mechanisms is physical barrier which classified to external physical barrier (skin, mucus and gills) and internal barrier represented by intestinal epithelial, the physical and chemical defenses are provided by these barriers (16; 5).

Skin act as first line defense mechanisms and classified as physical and chemical barrier for trapping pathogen (23), variable in structure according fish species in teleosts the skin have have scales known as Leptoid scals and other fish as cat fish lack scales the skin immune components of these types are skin-associated lymphoid tissue(SALT), secreting cells(gobalt cells,granulocytes,macrophage and langerhans -like cell and T lymphocyte(24)

as well as melanocytes in cartilaginous fish (25). Heimroth et al., (26) refer to present keratinocyte and B cells with granulocytes in skin of Lobe-finned fish, in general skin integrity and epidermal cells hyperplasia with these mononuclear cells and granulocyte is important to prevent entry of pathogen (27)

Gills

Addition to its biological function in keep gas exchanges and osmotic imbalance, gills conserved a physical barrier and having both innate and adaptive immunity which involve

macrophage and mononuclear cells and granulocytes (neutrophils and eosinophils) have been detected in gill associated lymphoid tissue (GALT), also B and T lymphocyte have been observed in some fish as in gill of rainbow trout, cat fish and shark in which B cells transcript to Igs (28; 29) also, expression of B form MHC II has been demonstrated in the gills of Salmonids (30).

Gastrointestinal (GI) tract

The gastrointestinal tract have biological function play roles in absorption of nutrient and also have both innate and adaptive immunity mainly posterior segment and diffuse cells (amyloid and lymphoid) are demonstrated in GALT which are not recognized as in high vertebrate, microflora have roles in development and maturation (31).

Mucus

One of the most important physical barriers is the mucus providing sub molecules which interact and inhibiting entry of pathogen (32). Mucus is more common secretion from skin and gills. In stress the stressor factors considered as stimulator for mucus secretion as microbial agent and chemical assembly, in some cases the mucus reach to 40% of the body weight in case Myxinoids (33). The mucus of fish composed from (i) Lectins are protein present also in eggs and blood their mechanisms is promoting agglutination because their binding to carbohydrate of microorganisms cells wall (ii) Lysozymes also, present in mucus which are peptide and can destroy membrane of pathogen (iii) Pentaxin (C-reactive protein) is inflammation phase protein has ability to connecting and promoting microorganisms opsonization, complement activation and phagocytosis, mucus have Ig and lysosomes which secreted from (monocytes and neutrophils) act on lysis peptidoglycan of microorganisms cell wall (34; 35).

Cellular Component

Fish possess different cell types as Natural killer cells (NK), dendritic cells, monocytes,

granulocyte (mast cells, eosinophilic and neutrophils), leukocyte are produce in head kidney and thymus in bony fish, while in cartilaginous fish the cells are produce in primary sites (epigonal organ, thymus, spleen and Leydig organ) (36; 37). Proliferation, survival, differentiation, maturation and biological function are regulated by cytokine act on cell receptors (38).

Toll-Like Receptors (TLRs)

TLRs have been distinguished by reported of (39) in *Carassius auratus auratus*, Pufferfish, zebra fish (40; 41) and in Japanese flounder (*Paralichthys olivaceus*) (42). These TLRs act for recognizing the molecules of the microbes called pathogen associated molecules patterns (PAMPs) this led to inflammation which is the first response of innate immunity (43).

Natural Killer Cells

T cytotoxic are nonspecific classified as innate immunity produced in lymphoid tissues a NK and spleen but rare found in blood (44), like NK in mammals they are small, granular lymphocyte invetidation in Catfish, Rainbow trout, Common carp and *Oreochromis spp.* (45; 46), their function was destroy virus and tumor cells (47; 48).

Dendritic cells

Several cells (thrombocyte) have ability to phagocytize pathogen and exogenous material as well as coagulation function, they have acid phosphatase lead to inflammation (49)

Macrophage is called monocyte in blood circulation and called macrophage in connective tissue, derived from hematopoietic progenitors. In general, is defined as transit cells which have ability to migrate during inflammation through tissue and turn to macrophage, have role in innate and adaptive immunity (50). They play roles in phagocytosis of pathogen, production of reactive oxygen species (ROS), nitric oxide (NO) as well as release pro-inflammation (TNF, IL-1B) these classified as

subpopulation as M_1 and have roles wound repair, immunosuppression and IL-10 M_2 (51), while in an adaptive immunity it known professional antigen presenting cell(pAPC) which act for presenting foreign (phagocytic) material to T lymphocyte (52)

The granulocyte cells in fish involve: neutrophils, polymorphonuclear cell, determined in blood, peritoneal cavity and lymphoid tissue, neutrophils play important roles in an innate defense against pathogen(53) that can phagocytosis pathogen or cells by producing extracellular and intracellular granules, they release reactive species, nitric oxide and myeloperoxidases in the cytoplasm which can kill bacteria halide with hydrogen peroxide and cause halogenation of the bacteria cell wall also, have lysosomal and other hydrolytic enzyme (54; 27).

Other granulocytes are eosinophils have acidophilic stain, large elongated homogenous granules found in connective tissue mainly in gill, GIT and blood stream, their function are degranulation and destroyed parasite (55)

Basophils are uncommon in fish, have a azurophilic grains in their cytoplasm, present in the blood circulation, their function unknown but through to provide and destroyed parasitic fish (56), in mammals have roles in allergy and antiparasitic immunity while in fish it activated by Ab-dependent manner in Fugu (*Takifugu rubripes*) distinguish IgM cross linked to basophils surface lead to degranulation of both two type reddish-purple and dark violet(57), some materials as papains stimulate degranulation of the reddish purple this will stimulate migration of other white blood cells while dark violet degranulation by chitin which stimulated series migration of CD4 T cells.

Other roles in adaptive immunity is interact with T and APCs and endocytosed Ag and expressed MHCII

Humeral and Innate Immunity

Anti-microbial peptide (AMPs)

It is known also as a host defense peptide, they composed of oligopeptide with different amino acid (cathelicidins, hepcidine, histone-derived peptide and specific piscidine), they demonstrated in gills, liver and mucus these AMPs have roles in innate immunity through destruction or forming pore against pathogen membrane (58), there are two type of AMPs (transferrin and squalamine) are distinguish in dog fish shark (*Squalus acanthias*) which act as bactericidal against gram positive and gram negative, other type known as AMPs Kenojenin has been distinguish in skin of fermented (*R. kenojei*) which act as inhibitory effects on *Bacillus subtilis*, *E. coli* and *Saccharomyces cerevisiae* (59; 60)

Lysosome

Is one of mediating defense mechanisms, is lytic enzyme cause hydrolyzing for peptidoglycan layer of bacteria (61). Lysosome are present in an invertebrate, plant, bacteriophage microbes, present in saliva, mucus and blood stream of high vertebrate while in fish they release from neutrophils and macrophage so it present on hematopoietic tissue mainly head kidney and in tissue exposed to pathogen invasion (gills, skin and gut tissue)(16).

There are two types of lysosomes distinguish in vertebrate goose (g)-type and chicken (c)-type, in fish mainly present both type in neutrophils and less extent in macrophage (62). In highly bacteriolytic activity against some pathogen as *Vibrio anguillarum*, *Aeromonas hydrophila* and *Micrococcus lysodeikticus* recombinant (r) between both g- and c- types will be occur (63).

Mode of action of lysosomes are attack lipopolysaccharide layer in pathogen cell all after earlier disruption of wall by complement and other enzyme lead to damage to enter structure, or cause disruption to wall permeability without lysis (32), so leukocyte released enzyme are more activity rather than

mammals (64).

Acute phase protein

It is a member of the pentraxin family present in egg, mucus and blood stream, Mantovani *et al.*, (65) identified it as first pattern recognition receptor (PRR), produce in both fish and mammals under factors release from fat cells and immune cells stimulate liver to produce acute phase protein (APP) (66), which have many activities as coagulation factors, transport protein (ferritin), serum amyloid protein (SAP) (67).

Also, another important acute phase protein is C-reactive protein (CRP) have ability to pathogen opsonization, complement and phagocytosis (35), some fish do not possess CRPs as Flounder, *Platichthys flesus* and the bacterial endotoxin (LPS) have ability to stimulate liver of fish exposed to pathogen to release CRP (68).

Complement

The complement system is cascade serum protein act in elimination of the pathogen through opsonization and phagocytosis and stimulate inflammation response, it is one of the main humeral components in both innate and adaptive immunity (69), in fish is considered effective than that mammals, but have same mode of action which can be activated by three ways (i) classical pathway triggered by Ab binding to cell wall of pathogen, is more common in mammals (70) (ii) alternative pathway which is dependent on Ab and activated directly by microorganisms or pAMPs this pathway more common in fish than in mammals (71) and finally (iii) lectin pathway in which interaction between lectin (is one component of complement) with sugar (mannose/mannan) present in the surface of pathogen (72).

Complement activate and neutralize envelop of virus while lipopolysaccharide in the cell wall of G⁻ stimulate C5a factor which is chemokines to macrophage and C3 was chemokines to neutrophils (73), the biological function of complement are cellular activation, chemotaxis, inflammation reaction

and phagocytosis and destroy pathogen through membrane injury and lysis (67)

Natural Antibodies

These types of Abs are present in the serum of fish when there have not infection or lack Ag stimulation of the cells that are equivalent to blood cells (74). They considered initiated key for innate immunity and linked to specific memory; teleost fish have IgM neutral Ab provide protection against different Ags. Species of fish and variable environmental condition represented commonly factors affecting generation and levels of natural Abs (75).

Adaptive Immunity

The adaptive refer to recognition of specific form from non-specific and specialized responded to pathogen if persist and surviving innate immunity which activated adaptive immunity (76). The main components of humeral are Abs and cytotoxic T-lymphocyte cells which representing and mainly component of cellular adaptive immunity (77), humeral immunity similar to humeral high vertebrate by Igs structure, function (neutrolization, complement fixation and opsonization also, the cellular which responsible for Abs stimulation but in fish affected by fish species and environment factors.

Antibodies

Known also immunoglobulin Ig, are major component and primary humeral immunity (10), teleost Abs are found systematically in plasma or may found locally in gills, intestine, bile and mucus (78), gills are more organ direct contact to environment pathogen and stressors factors so the immune response and defense mechanisms are important (79), their function may be explaining as Ag receptors on the surface of b cells or Abs secreted to blood (80).

Types of teleost Abs:

(1) IgM: commonly and predominant Ig in all fish as tetrameric and monomeric, have eight sites for antigenic combining exception that

coelacanth do not IgM in the genome (81) also, in cartilaginous fish IgM found pentameric and monomeric which play roles in phagocytosis (82)

In bony fish and cartilaginous fish IgM are present in two forms :transmembrane which is shorter than second form which is secreted led to shorter IgM receptors in B cells but these do not affected their function in both innate and adaptive immunity which are Ab-dependent cell- mediated cytotoxicity, opsonization and complement activation (83) .

(2) IgD/IgW:IgD is present in all bon fish, while IgW found in cartilaginous fish(84), coelacanths and lung fish (85)both Igs are phylogenetically old as IgM (86).

IgD have play roles in innat immunity secreted by channel catfish (87) mainly in gill, (88) suggested that IgD /IgM ration in gill much higher than other tissue, also IgN -/IgD + B cell are expressed in the gill.

(3) SPECIEFIC Ab related to species:

IgS:IgNAR, IgZ/T ,IgQ IgZ is firstly identified inzebrafish (89) and IgT in rainbow trout , it specialized for for mucosal immunity and functionally an alogously to mammams IgA.

The IgT/Z in the serum lower than IgM, in general the ration IgT/Z to IgM higher in the gut than serum (90) IgT +B cell also detecated in fish skin and lymphoid tissue (91)

IgNAR (new/nurse shark antigen receptor) is a heavy chain Ig found in shark, in the serum levels of IgNAR much lower than IgM (92).

Immunological Memory (B cells)

B cells are lymphocytes have avital functions which is responsible for produce Ab, selected as memory B cells which have greater and higher affinity receptors in status of memory response which has faster and larger than primary responsealso the number of Ag-specific B cells in the spleen proportional to B cells – specific Ag (93; 78) also, B cells act as APCs, teleost fish have been lack bone marrow so the main hematopoietic for progenitor B cell and plasma cell is anterior kidney(7) while posterior kidney and spleen representing sites for maturation and activation B cell and plasma blast formation

(94;95) and then differentiation in to plasma cells ,these cells migration back to the anterior where have long life span and supporting the storage of larger amount of Ig secreting (96).

Also, B cells found in other organs as in skin of catfish (97), primary gill filaments a long blood vessel in Spotted wolfish (98), in the intestine of carp, sea bass and brain bow trout (99).

T-cells and Cellular cytotoxicity

Lymphocyte mature in to the thymus so it called thymocyte, there are many types involve gamma delta T cell, regulatory Tcell (Treg cell), memory T cell, T helper cells (T_H cell) and T cytotoxic cell (CTL) (100; 101).

CD+8 cytotoxic and CD+3 have ability to recognize and kill virus with derived peptides using MHCI (27), CD+4 T cells have plasticity and heterogeneity (102).There are five types of naive CD+4T cell can recognize to effectors cells (Th1,Th9,Th2,Th22 and Th17). Fischer *et al.*, (103) suggested that CD+4 helper and CD+4 CTL in fish similar to higher invertebrate.The presence of T cells is detected at variable time according to fish species As in Sea bass fish (*Dicentrarchus labrax*) T cell was distinguishing in larval stage at five days after incubation (14) and after one week of rainbow trout insemination the CD8+T cells have been distinguish

Cytokines

As modulators of the adaptive immune response there was a little data about teleost cytokines which include:

Interleukine -1B

It detected in both bony and cartilaginous fish, produce by macrophage it is play vital roles as mediator of inflammation and stimulating cortisol secretion (104), other cytokines and regulatory molecules reported in fish with interferon(IFN-g) which are effectors of the Th1 response as(IL-12, IL-18, IL-15)(105; 106; 107) and IL-10 has effectors of T regulatory cells (108)while IL-4 has been relation to Th2 type(109).

Tumor Necrosis Factors (TNF)

TNF- alpha and TNF-beta have been cloned

in all fish species rainbow trout, catfish, *Carassius auratus* and sea bream(*Sparus aurata*) play an important roles in neutrophils migration, phagocytosis and burst activity of macrophage and nitric oxide production as well as induce apoptosis and considered a key factor for neuroimmunoendocrine responses (110; 111; 112).

Interferon (IFN)

There are two types of these cytokines IFN-alpha and INF-beta, both have antiviral activity by inhibition viral nucleic acid replication,also play another role in protection cells from viral infection by binding to variable receptors lead to induction several genes that are stimulated by INF(ISGs), these genes are encodes antiviral protein as(MX) (113 and 114).

Interferon like peptide has been detection in zebra fish (115), IFN- alpha -1 was recognized in *Atlantic salmon* induces expression of MX and ISGs and with IFN-alpha-2 have activity against infectious pancreatic necrosis virus (116 and 117).

Conclusion

The immune system of fish is sufficient as in mammalian to provide defense against antagonistic pathogens from the environment and improving the safety of the health and disease of fish.

References

- 1-Ausubel FM. Are innate immune signaling pathways in plants and animals conserved? *Nat. Immunol.* 2005 Oct;10: 973–979. DOI: 10.1038/ni1253
- 2-Janeway CA and Medzhitov R. Innate immune recognition. *Annual Review of Immunology.*2002 Oct; 20:197- 216. Jang doi: 10.1146/annurev.immunol.20.083001.084359
- 3- David D, Chaplin, MD. Overview of the Immune Response. *The Journal of allergy and clinical immunology.* 2010 Feb; 125(2 Suppl 2), S3-23. <http://dx.doi.org/10.1016/j.jaci.2009.12.980>
- 4-Van Muiswinkel WB. The Piscine Immune System: Innate and Acquired Immunity.Chapter 17 in”Fish Disorders 1:Protozoan and Metazon Infections”, P.T.K.Woo(Editor),CAB International,Wallingford,UK, 1995:725-750. <https://www.researchgate.net/publication/40207091>
- 5-Tort L, Balasch JC, Mackenzie S .Fish Immune System.A crossroads between innate and adaptive responses. *Immunologica.* 2003 Sep; 22(3):277-286. <https://www.immunologia.org/Upload/Articles/6/0/602.pdf>
- 6-Salinas I, Zhang YA, Sunyer JO. Mucosal immunoglobulins and B cells of teleost fish. *Dev Comp Immuno.* 2011 Dec; 35: 1346-1365. doi: 10.1016/j.dci.2011.11.009.
- 7-Zapata A, Diez B, Cejalvo T, Gutierrez-De Frias C, Cortes A. Ontogeny of the immune system of fish. *Fish and Shellfish Immunology.*2006 Feb; 20, 126–136 doi: 10.1016/j.fsi.2004.09.005.
- 8-Mulero V, Meseguer J. Functional charaterisation of a macrophage -activating factor produced by leucocytes of gilthead seabream (*Sparus aurata L.*). *Fish and Shellfish Immunology.*1998. 8,143-156.
- 9-Willett C, Cortes, A Zuasti A,Zapata A. Early hematopoiesis and developing lymphoid organs in the zebrafish. *Developmental Dynamics .*1999 Apr; 214, 323–336. DOI: 10.1002/(SICI)1097-0177(199904)214:4<323: AID-AJA5>3.0.CO;2-3

- 10-Magnadottir B, Lange S, Gudmundsdottir S, Bogwald J, Dalmo RA. Ontogeny of humoral immune parameters in fish. *Fish and Shellfish Immunology*. 2005 Nov; 19: 429–439. DOI: 10.1016/j.fsi.2005.03.010
- 11-Dalmo RA. Ontogeny of the fish immune system. *Fish and Shellfish Immunology*. 2005 Nov; 19:395–396. DOI: 10.1016/j.fsi.2005.03.004.
- 12-Ferguson HW. *Systemic Pathology of Fish. A text and atlas comparative tissue response in diseases of teleost*. Iowa state University Press. Ames. Iowa, USA. 1998, 5–103.
- <https://www.amazon.com/Systemic-Pathology-Fish-Comparative-Responses/dp/0813801478>.
- 13-Solem ST , Stenvik J. Antibody repertoire development in teleosts—a review with emphasis on salmonids and *Gadus morhua* L. *Dev Comp Immunol*. 2006 Jul; 30: 57-76. doi: 10.1016/j.dci.2005.06.007.
- 14-Dos Santos NM, Romano N, de Sousa M, Ellis AE, Rombout JH. Ontogeny of B and T cells in sea bass (*Dicentrarchus labrax*, L.). *Fish and Shellfish Immunology*, 2000 Oct; 10: 583–596
- 15-Petrie-Hanson L, Ainsworth AJ. Differential cytochemical staining characteristics of channel catfish leukocytes identify cell populations in lymphoid organs. *Veterinary Immunology and Immunopathology*. 2000 March; 73: 129–144.
- 16-Ellis AE. Innate host defence mechanism of fish against viruses and bacteria. *Developmental and Comparative Immunology*. 2001 Oct-Dec; 25: 827–839. doi: 10.1016/s0145-305x(01)00038-6.
- 17-Davis JM, Clay H, Lewis JL, Ghori N, Herbomel Ramakrishnan L. Real-time visualization of mycobacterium-macrophage interactions leading initiation of granuloma formation in zebrafish embryos. *Immunity*. 2002 Dec; 17: 693–702 doi: 10.1016/s1074-7613(02)00475-2
- 18-Torroba M, Zapata AG. Aging of the vertebrate immune system. *Microsc Res Technol*. 2003 Nov; 62: 477-481. doi:10.1002/jemt.10409
- 19-Silva-Gomes S, Decout, A, Nigou J. Pathogen Associated Molecular Patterns (PAMPs). In M. Parnham (Ed.), *Encyclopedia of Inflammatory Diseases*. 2015; 1-16. Basel: Springer Basel.
- 20-Taghavi M, Khosravi A, Mortaz E, Nikaein D و Athari SS. Role of pathogen-associated molecular patterns (PAMPs) in immune responses to fungal infections. *European Journal of Pharmacology*. 2017 Nov; 808: 8-13. <http://dx.doi.org/10.1016/j.ejphar.2016.11.013>.
- 21-Alberts B, Johnson A, Lewis J, Raff M, Roberts K, Walter P. Helper T cells and lymphocyte activation. In: *Molecular Biology of the Cell*. 4th Edn. New York, NY: Garland Science). 2002. Available Online at: <https://www.ncbi.nlm.nih.gov/books/NBK26827/> ISBN-10: 0-8153-3218-1.
- 22-Smith NC, Rise ML, Christian S. A Comparison of the Innate and Adaptive Immune System in Cartilaginous Fish, Ray – Finned Fish, and Lobe- Finned Fish. *Frontiers in Immunology*. 2019 Oct; 10. DOI: [org/10.3389/fimmu.2019.02292](https://doi.org/10.3389/fimmu.2019.02292).
- 23-Esteban MÁ. An overview of the immunological defenses in fish skin. *ISRN Immunol*. 2012 Oct; 29 pages. doi: 10.5402/2012/853470.

- 24-Xu Z, Parra D, Gomez D, Salinas I, Zhang YA, von Gersdorff Jorgensen L, Heinecke RD, Buchmann K, LaPatra S, Sunyer JO. Teleost skin, an ancient mucosal surface that elicits gut-like immune responses. *Proc Natl Acad Sci USA*. 2013 Aug; 110(32):13097–102. doi: 10.1073/pnas.1304319110.
- 25-Meyer W, Seegers U. Basics of skin structure and function in elasmobranchs: a review. *J Fish Biol*. 2012 Apr; 80(5):1940–67. doi: 10.1111/j.1095-8649.2011.03207.x
- 26-Heimroth RD, Casadei E, Salinas I. Effects of experimental terrestrialization on the skin mucus proteome of African lungfish (*Protopterus dolloi*). *Front Immunol*. 2018 Jun; 9:1259. doi: 10.3389/fimmu.2018.01259.
- 27-Fischer U, Utke K, Somamoto T, Kollner B, Ototake M, Nakanishi T. Cytotoxic activities of fish leucocytes. *Fish and Shellfish Immunology*. 2006 Feb; 20(2): 209–226. doi: 10.1016/j.fsi.2005.03.013.
- 28-Criscitiello MF, Ohta Y, Saltis M, McKinney EC, Flajnik MF. Evolutionarily conserved TCR binding sites, identification of T cells in primary lymphoid tissues, and surprising trans-rearrangements in nurse shark. *J Immunol*. 2010 May; 184:6950–60. doi: 10.4049/jimmunol.0902774
- 29-Ortiz NN, Gerdol M, Stocchi V, Marozzi C, Randelli E, Bernini C, Buonocore F, Picchiatti S, Papeschi C, Sood N, Pallavicini A, Scapigliati G. T cell transcripts and T cell activities in the gills of the teleost fish sea bass (*Dicentrarchus labrax*). *Dev Comp Immunol*. 2014 Dec; 47:309–18. doi: 10.1016/j.dci.2014.07.015.
- 30-Picchiatti S, Terribili FR, Mastrolia L, Scapigliati G, Abelli L. Expression of lymphocyte antigenic determinants in developing gut-associated lymphoid tissue of the sea bass *Dicentrarchus labrax* (L.). *Anat Embryol*. 1998 Jan; 196(6):457–63. doi: 10.1007/s004290050113.
- 31-Wang AR, Ran C, Ring Ñ E, Zhou ZG. Progress in fish gastrointestinal microbiota research. *Rev Aquacult*. 2018 Feb; 10:626–40. doi: 10.1111/raq.12191.
- 32-Saurabh S, Sahoo PK. Lysozyme: an important defense molecule of fish innate immune system. *Aquaculture Research*. 2008 Jan; 39:223–239. DOI: 10.1111/j.1365-2109.2007.01883.x
- 33-Hooper LV, Littman DR and Macpherson AJ. Interactions between the microbiota and the immune system. *Science*. 2012 Jun; 336:1268–73. doi: 10.1126/science.1223490.
- 34-Goldsby RA, Kindt TJ, Osborne BA, Kuby J. In: *Immunology*, 5th ed., WH Freeman & Company Publisher. 2002. 554 p.
- 35-Magnadóttir B, Audunsdóttir SS, Bragason B Th, Gísladóttir B, Jónsson ZO, Gudmundsdóttir S. The acute phase response of Atlantic cod (*Gadus morhua*): Humoral and cellular responses. *Fish Shellfish Immunol*. 2011 Apr-May; 30: 1124-1130. doi: 10.1016/j.fsi.2011.02.010.
- 36-Zardakis IK, Mastellos D, Lambris JD. Phylogenetic aspects of the complement system. *Dev Comp Immunol*. 2001 Oct-Dec 2001; 25(8-9):745-62. DOI: 10.1016/s0145-305x(01)00034-9.
- 37-Chettri JK, Raida MK, Andersen LH, Kania PW, Buchmann K. PAMP induced expression of immune relevant genes in head kidney leukocytes of rainbow trout (*Oncorhynchus mykiss*). *Dev Comp Immunol*. 2011 Dec; 35: 476-482. DOI: 10.1016/j.dci.2010.12.001.
- 38-Hanington PC, Tamb J, Katzenback BA, Hitchen SJ, Barreda DR, Belosevic M.

- Development of macrophages of cyprinid fish. *Dev Comp Immunol.* Jan; 2009 33: 411-429. DOI: [org/10.1590/0001-3765201420130159](https://doi.org/10.1590/0001-3765201420130159).
- 39-Stafford JL, Ellestad KK, Magor KE, Belosevic M, Magor BG. A toll like receptor (TLR) gene that is up-regulated in activated goldfish macrophages. *Developmental Comp.Immunol.* 2003 Sep; 75: 685-698.doi: 10.1016/s0145-305x (03)00041-7
- 40-Oshiumi H, Tsujita T, Shida K, Matsumoto M, Ikeo K, Seya T. Prediction of the prototype of the human Toll-like receptor gene family from the pufferfish, *Fugu rubripes*, genome. *Immunogenetics.* 2003 Feb; 54(11): 791–800.doi: 10.1007/s00251-002-0519-8.
- 41-Jault C, Pichon, L, Chluba J.Toll-like receptor gene family and TIR-domain adapters in *Danio rerio*. *Molecular Immunology.*2004 Jan; 40(11), 759–771.
- 42-Takano T, Kondo H, Hirono I, Endo M, SaitoTaki T, Ao T. Toll-like receptors in teleosts. *Diseases in Asian Aquaculture VII Fish Health Section, Asian Fisheries Society, Malaysia.* 2011. 197-208.
- 43-Akira S, Uematsu S, Takeuchi O.Pathogen recognition and innate immunity. *Cell,*2006 Feb; 124(4), 783–801. doi: 10.1016/j.cell.2006.02.015
- 44-Shen L, Stuge TB, Zhou H, Khayat M, Barker KS, Quiniou SM, Wilson M, Bengten E, Chinchar VG, Clem LW, Miller NW .Channel catfish cytotoxic cells: a mini-review. *Developmental and Comparative Immunology.*2002 Mar; 26(2): 141–149. DOI: 10.1016/s0145-305x (01)00056-8.
- 45-Greenlee AD, Brown RA, Ristlow SS. Nonspecific cytotoxic cells of Rainbow trout (*Oncorhynchus mykiss*) kill Yac-1 targets by both necrotic and apoptotic mechanisms. *Developmental and Comparative Immunology.* 1991 Jan; 15(3):153–164.DOI:10.1016/0145-305X (91)90006-K
- 46-Suzumura E, Kurata O, Okamoto N, Ykeda Y. Characteristics of natural killer cells in carp. *Fish Pathology.* 1994 May; 29(3):199–203.
https://www.jstage.jst.go.jp/article/jsfp1966/29/3/29_3_199/
- 47-Tizard IR. *Imunologia Veterinária. Uma introdução.* Editora Roca, São Paulo. 2002.532 p.
- 48-Raulet DH. Interplay of natural killer cells and their receptors with the adaptive immune response. *Nature Immunol.* 2004 Oct; 5(10): 996-1002. doi: 10.1038/ni1114.
- 49-Tavares-Dias M, Schalch SHC, Martins ML, Silva ED, Moraes FR, Perecin D . Hematologia de teleósteos brasileiros com infecção parasitaria. I. Variáveis do *Leporinus macrocephalus* Garavello & Britski, 1988 (Anostomidae) e *Piaractus mesopotamicus* Holmberg, 1887 (Characidae). *Acta Sci Anim Sci.* 1999 July; 21(2): 337-342.
- 50-Cuesta A, Angeles esteban M, Meseguer J. Natural cytotoxic activity of gilthead seabream (*Sparus aurata* L.) leucocytes assessment by flow cytometry and microscopy. *Vet Immunology Immunopath.* 1999 Nov; 71: 161-171.
- 51-Joerink M, Ribeiro CM, Stet RJ, Hermsen T, Savelkoul HF, Wiegertjes GF. Head kidney-derived macrophages of common carp (*Cyprinus carpio* L.) show plasticity and functional polarization upon differential stimulation. *J Immunol.* 2006 July; 177:61–9. doi: 10.4049/jimmunol.177.1.61.
- 52-Hodgkinson JW, Grayfer L, Belosevic M.Biology of bony fish macrophages. *Biology.* 2015 Nov 4(4):881–906. doi: 10.3390/biology4040881.
- 53-Havixbeck JJ, Barreda DR. Neutrophil

- development, migration, and function in teleost fish. *Biology*. 2015 Nov 4(4):715–34. doi: 10.3390/biology4 040715. doi: 10.3390/biology4040715.
- 54-Neumann NF, Stafford JL, Barreda D, Ainsworth AJ , Belosevic M. Antimicrobial mechanisms of fish phagocytes and their role in host defense. *Dev Comp Immunol*. 2001 Oct-Dec 2001;25(8-9):807-25. doi: 10.1016/S0145-305X (01)00037-4
- 55-Hine PM. The granulocytes of fish. *Fish Shellfish Immunol*. 1992 Apr; 2: 79-88.
- 56-Martins ML, Moraes FR, Moraes JRE, Malheiros EB . Falha na resposta do cortisol estresse por captura e por carreginina em *Piaractus mesopotamicus* Holmberg (Osteichthyes, Characidae). *Rev Bras Zool*. 2000. 12: 545-552.
- 57-Kirsten K., Fior D, Kreutz LC and Barcellos LJG. First description of behavior and immune system relationship in fsh. *Sci Rep*. 2018 Jan; 8:846. www.nature.com/scientificreports, DOI:10.1038/s41598-018-19276-3
- 58-Birkemo GA, Luders T, Andersen O, Nes IF, NissenMeyer J. Hippusin, a histone-derived antimicrobial peptide in Atlantic halibut (*Hippoglossus hippoglossus* L.). *Journal of Biochemistr*. 2003 Mar 21;1646(1-2):207-15.DOI: 10.1016/s1570-9639(03)00018-9.
- 59-Bates JM, Mittge E, Kuhlman J, Baden KN, Cheesman SE, Guillemin K. Distinct signals from the microbiota promote different aspects of zebrafish gut differentiation. *Dev Biol*. 2006 Sep; 15;297(2):374-86.doi: 10.1016/j.ydbio.2006.05.006
- 60-Choe SH, Lee BD, An H, Eun JB, Kenojein I. Antimicrobial peptide isolated from the skin of the fermented skate, *Raja kenoei*. *Peptides*. 2005 April; 26:581–7. doi: 10.1016/j.peptides.2004.11.011.
- 61-Brazeau MD and Friedman M. The origin and early phylogenetic history of jawed vertebrates. *Nature*. April 23;520(7548):490-7.doi: 10.1038/nature14438.
- 62-Myrnes B, Seppola M, Johansen A, Overbo K, Callewaert L, Vanderkelen L, Michiels CW ,Nilsen IW.lysozymes. *Dev Comp Immunol*. 2013 May;40(1):11-9. doi: 10.1016/j.dci.2013.01.010.
- 63-Buonocore F, Randelli E, Trisolino P, Facchiano A, de Pascale D, Scapigliati G. Molecular characterization, gene structure and antibacterial activity of a g-type lysozyme from the European sea bass (*Dicentrarchus labrax* L.). *Mol Immunol*. 2014 Nob; 62(1):10–8. doi: 10.1016/j.molimm.2014.05.009.
- 64-Demers NE, Bayne CJ. The immediate effects of stress on hormones and plasma lysozyme in rainbow trout. *Developmental and Comparative Immunology*.Jul-Aug 1997; 21(4):363-73. doi: 10.1016/s0145-305x(97)00009-8.
- 65-Mantovani A, Garlanda C, Doni A, Bottazzi B. Pentraxins in innate immunity: From C-reactive protein to the long pentraxin PTX3. *Journal of Clinical Immunology*. 2008 Jan; 28(1): 1–13. doi: 10.1007/s10875-007-9126-7.
- 66-Roy S, Kumar V, Kumar V, Behera BK (2017). Acute phase proteins and their potential role as an indicator for fish health and in diagnosis of fish diseases. *Protein Pept Lett*, 24:78–89. doi: 10.2174/0929866524666161121142221.
- 67-Bayne CJ, Gerwick L.The acute phase response and innate immunity of fish. *Dev*

- Comp Immunol. 2001 Oct-Dec; 25(8-9):725-43. doi: 10.1016/S0145-305X(01)00033-7.
- 68-White A, Fletcher TC. The influence of hormones and inflammatory agents on C-reactive protein, cortisol, and alanine aminotransferase in the plaice (*Pleuronectes platessa L.*). *Comparative Biochemistry and Physiology.* 1985 Jan; 80(1): 99–104. doi: [10.1016/0742-8413\(85\)90138-0](https://doi.org/10.1016/0742-8413(85)90138-0)
- 69-Nakao M, Tsujikura M, Ichiki SK, Vo T, Somamoto T. The complement system in teleost fish: Progress of post-homolog-hunting researches. *Developmental & Comparative Immunology.* 2011 March; 35(12): 1296–1308. doi: [10.1016/j.dci.2011.03.003](https://doi.org/10.1016/j.dci.2011.03.003)
- 70-Gasque PH. Complement: A unique innate immune sensor for danger signals. *Molecular Immunology.* 2004 Nov; 41(11): 1089–1098. doi: [10.1016/j.molimm.2004.06.011](https://doi.org/10.1016/j.molimm.2004.06.011).
- 71-Yano T. The nonspecific immune system Humoral defense. In Hoar, W. S., Randall D. J., Iwama, G., & Nakanishi, T. (Eds.), *The fish immune system: Organism, pathogen and environment.* Fish immunology series. New York: Academic Press. 1996
- 72-Fujita T, Matsushita M, Endo Y. The lectin-complement pathway – Its role in innate immunity and evolution. *Immunology Review.* 2004 Apr; 198:185-202. doi: [10.1111/j.0105-2896.2004.0123.x](https://doi.org/10.1111/j.0105-2896.2004.0123.x).
- 73-Lorenzen N, LaPatra SE. Immunity to rhabdovirus in rainbow trout: the antibody response. *Fish and Shellfish Immunology.* 1999 May; 9: 345–360. <https://doi.org/10.1006/fsim.1999.0194>
- 74-Boes M. Role of natural and immune IgM antibodies in immune responses. *Molecular Immunology.* 2000 Dec; 37(18):1141-9. doi: [10.1016/s0161-5890\(01\)00025-6](https://doi.org/10.1016/s0161-5890(01)00025-6)
- 75-Whyte SK. The innate immune response of finfish e A review of current knowledge. *Fish and Shellfish Immunology.* 2007 Jan; 23: 1127–1151. doi: [10.1016/j.fsi.2007.06.005](https://doi.org/10.1016/j.fsi.2007.06.005)
- 76-Flajnik MF. A cold-blooded view of adaptive immunity. *Nat Rev Immunol.* 2018 Jul; 18(7):438–53. doi: [10.1038/s41577-018-0003-9](https://doi.org/10.1038/s41577-018-0003-9).
- 77-Firdaus-Nawi M, Zamri-Saad M. Major Components of Fish Immunity: A Review. *Pertanika J. Trop. Agric. Sci.* 2016 Sep; 39(4): 393 – 420. Journal homepage: <http://www.pertanika.upm.edu.my>
- 78-Morrison RN, Nowak BF. The antibody response of teleost fish. *Seminars in Avian and Exotic Pet Medicine.* 2002 Jan; 11(1): 46–54. <https://doi.org/10.1053/saep.2002.28241>
- 79-Cain KD, Jones DR, Raison RL. Characterisation of mucosal and systemic immune responses in rainbow trout (*Oncorhynchus mykiss*) using surface plasmon resonance. *Fish and Shellfish Immunology.* 2000 Nov; 10(8): 651–666.
- 80-Pilstrom L, Bengten E. Immunoglobulin in fish – Genes, expression and structure. *Fish and Shellfish Immunology.* 1996 May; 6(4):243–262. <https://doi.org/10.1006/fsim.1996.0026>.
- 81-Saha NR, Ota T, Litman GW, Hansen J, Parra Z, Hsu E, Buonocore F, Canapa A, Cheng JF, Amemiya CT. Genome complexity in the coelacanth is reflected in its adaptive immune system. *J Exp Zool B Mol Dev Evol.* 2014 Jan; 322:438–63. doi: [10.1002/jez.b.22558](https://doi.org/10.1002/jez.b.22558).
- 82-Dooley H, Flajnik MF. Shark immunity bites back: affinity maturation and memory response in the nurse shark, *Ginglymostoma cirratum*. *Eur J Immunol.* 2005 Mar; 35(3):936-45. doi: [10.1002/eji.200425760](https://doi.org/10.1002/eji.200425760)

- 83-Boshra H, Gelman AE, Sunyer JO. Structural and functional characterization of complement C4 and C1s-like molecules in teleost fish: insights into the evolution of classical and alternative pathways. *J Immunol*, 2004 August; 173(1):349–59. doi: 10.4049/jimmunol.173.1.349.
- 84-Anderson MK, Strong SJ, Litman RT, Luer CA, Amemiya CT, Rast JP, Litman GW. A long form of the skate IgX gene exhibits a striking resemblance to the new shark IgW and IgNARC genes. *Immunogenetics*. 1999 Jan; 49:56–67. doi: 10.1007/s002510050463.
- 85-Ota T, Rast JP, Litman GW, Amemiya CT. Lineage-restricted retention of a primitive immunoglobulin heavy chain isotype within the Dipnoi reveals an evolutionary paradox. *Proc Natl Acad Sci USA*. 2003 Mar; 100:2501–6. doi: 10.1073/pnas.0538029100.
- 86-Zhu L, Yan Z, Feng M, Peng D, Guo Y, Hu X, Ren L, Sun Y. Identification of sturgeon IgD bridges the evolutionary gap between elasmobranchs and teleosts. *Dev Comp Immunol*. 2014 Feb; 42:138–47. doi: 10.1016/j.dci.2013.08.020
- 87-Edholm ES, Bengten E, Stafford JL, Sahoo M, Taylor EB, Miller WN and Wilson M (2010). Identification of two IgD⁺ B cell populations in channel catfish, *Ictalurus punctatus*. *Journal of Immunology*, 185(7): 4082–4094.
- 88-Parra D, Korytar T, Takizawa F, Sunyer JO. B cells and their role in the teleost gut. *Dev Comp Immunol*. Mar; 64:150–66. doi: 10.1016/j.dci.2016.03.013.
- 89-Danilova N, Bussmann J, Jekosch K, Steiner LA. The immunoglobulin heavy-chain locus in zebrafish: identification and expression of a previously unknown isotype, immunoglobulin Z. *Nat Immunol*. 2005 Mar; 6(3):295–302. doi: 10.1038/ni1166.
- 90-Zhang YA, Salinas I, Li J, Parra D, Bjork S, Xu Z, LaPatra SE, Bartholomew J, Sunyer JO. IgT, a primitive immunoglobulin class specialized in mucosal immunity. *Nat Immunol*. 2010 Sep; 11(9):827–35. doi: 10.1038/ni.1913.
- 91-Xu Z, Parra D, Gomez D, Salinas I, Zhang YA, Jorgensen L VF, Heinecke RD, Buchmann K, LaPatra S, Sunyer JO. Teleost skin, an ancient mucosal surface that elicits gut-like immune responses. *Proc Natl Acad Sci USA*. 2013 August; 110 (32):13097–102. doi: 10.1073/pnas.1304319110.
- 92-Pettinello R, Dooley H. The immunoglobulins of cold-blooded vertebrates. *Biomolecules*. 2014 Nov; 4(4):1045–69. doi: 10.3390/biom4041045.
- 93-Kaattari SL. Fish B lymphocytes: Defining their form and function. In: Faisal M, Hetrick FM (eds.): *Annual Review of Fish Diseases*. Vol. 2. Pergamon, Tarrytown, NY, USA. 1992. 161–180.
- 94-Zwollo P, Cole S, Bromage E, Kaattari S. B cell heterogeneity in the teleost kidney: evidence for a maturation gradient from anterior to posterior kidney. *J Immunol*. 2005 Jun, 174(11):6608–16. doi: 10.4049/jimmunol.174.11.6608
- 95- Zwollo P. Dissecting teleost B cell differentiation using transcription factors. *Dev Comp Immunol*. 2011 Sep 35(9):898–905. doi: 10.1016/j.dci.2011.01.009.
- 96-Bromage ES, Kaattari IM, Zwollo P, Kaattari SL. Plasmablast and plasma cell production and distribution in trout immune tissues. *J Immunol*. 2004 Dec; 173(12):7317–23. doi: 10.4049/jimmunol.173.12.7317.
- 97-Zhao X, Findly RC, Dickerson HW. Cutaneous antibody-secreting cells and B cells in a teleost fish. *Developmental and*

- Comparative Immunology. 2008 August; 32(5): 500–508.
- 98-Grøntvedt RN, Espelid S. Vaccination and immune responses against atypical *Aeromonas salmonicida* in spotted wolffish (*Anarhichas minor* Olafsen) juveniles. *Fish and Shellfish Immunology*. 2004 Mar; 16(3):271–285.
- 99-Salinas I Zhang YA, Sunyer JO. Mucosal immunoglobulins and B cells of teleost fish. *Developmental and Comparative Immunology*. 2011 Nov; 35(12): 1346–1365.
- 100-Annunziato F, Romagnani S. Heterogeneity of human effector CD4+ T cells. *Arthritis Research and Therapy*. 2009 Dec; 11(6): 257. doi: 10.1186/ar2843.
- 101-Wan Y, Flavell RA. How diverse – CD4 effector T-cells and their functions. *Journal of Molecular Cell Biology*. 2009 May; 1(1): 20–36. doi: 10.1093/jmcb/mjp001.
- 102-Zhu J, Paul WE. Peripheral CD4+ T-cell differentiation regulated by networks of cytokines and transcription factors. *Immunology Review*. 2010 Nov; 238(1): 247–22. doi: 10.1111/j.1600-065X.2010.00951.x.
- 103-Fischer U, Koppang EO, Nakanishi T. Teleost T and NK cell immunity. *Fish Shellfish Immunology*. 2013 August; 35(2): 197–206. doi: 10.1016/j.fsi.2013.04.018.
- 104-Holland JW, Pottinger TG, Secombes CJ. Recombinant interleukin1 beta activates the hypothalamic-pituitary-interrenal axis in rainbow trout, *Oncorhynchus mykiss*. *J Endocrinol*. 2002 Oct; 175: 261267. DOI: 10.1677/joe.0.1750261.
- 105-Nascimento DS, do Vale A, Toma's AM, Zou J, Secombes CJ, dos Santos NMS (2007): Cloning, promoter analysis and expression in response to bacterial exposure of sea bass (*Dicentrarchus labrax* L.) interleukin-12 p40 and p35 subunits. *Molecular Immunology* 44, 2277–2791. doi: 10.1016/j.molimm.2006.11.006.
- 106-Zou J, Bird S, Truckle J, Bols N, Horne M, Secombes CJ. Identification and expression analysis of an IL-18 homologue and its alternatively spliced form in rainbow trout *Oncorhynchus mykiss*. *European Journal of Biochemistry*. 2004 Apr; 271:1913–1923. DOI: 10.1111/j.1432-1033.2004.04101.x.
- 107-Wang T, Holland J, Bols N, Secombes CJ. Molecular and functional characterisation of interleukin-15 in rainbow trout *Oncorhynchus mykiss*: a potent inducer of interferon-gamma expression in spleen leucocytes. *Journal of Immunology*. 2007 August; 179:1475–1488. DOI: <https://doi.org/10.4049/jimmunol.179.3.1475>.
- 108-Hardie LJ, Laing KJ, Daniels GD, Grabowski PS, Cunningham C, Secombes CJ. Isolation of the first piscine transforming growth factor β gene: analysis reveals tissue specific expression and a potential regulatory sequence in rainbow trout (*Oncorhynchus mykiss*). *Cytokine*. 1998 August; 10, 555–563. doi: 10.1006/cyto.1997.0334.
- 109-Li JH, Shao JZ, Xiang LX, Wen Y. Cloning, characterization and expression analysis of pufferfish interleukin-4 cDNA: the first evidence of Th2-type cytokine in fish. *Molecular Immunology*. 2007 Mar; 44, 2078–2086. DOI: 10.1016/j.molimm.2006.09.010.
- 110-Bobe J, Goetz FW. Molecular cloning and expression of a TNF receptor and two TNF ligands in the fish ovary. *Comp Biochem Physiol B Biochem Mol Biol*. 2001 Jun; 129: 475–481. DOI: 10.1016/s1096-4959(01)00353-0.
- 111-Qin QW, Ototake M, Noguchi K, Soma GI, Yokomizo Y, Nakanishi T. Tumor

necrosis factor alpha (TNFalpha)-like factor produced by macrophages in rainbow trout, *Oncorhynchus mykiss*. *Fish Shellfish Immunol.* 2001 Apr; 11: 245-256. <https://doi.org/10.1006/fsim.2000.0311>.

112- Tafalla C, Figueras A, Novoa B. Viral hemorrhagic septicemia virus alters turbot *Scophthalmus maximus* macrophage nitric oxide production. *Disease of Aquatic Organism.* 2001 Nov; 47(2): 101–107. doi: 10.3354/dao047101

113- Robertsen B. The interferon system of teleost fish. *Fish and Shellfish Immunology.* 2006 Feb; 20(2):172–191. doi: 10.1016/j.fsi.2005.01.010.

114- Samuel CE. Antiviral actions of interferons. *Clinical Microbiology Review.* 2001 Oct; 14:778–809. doi: 10.1128/CMR.14.4.778-809.2001.

115- Altmann SM, Mellon MT, Distel DL, Kim CH. Molecular and Functional Analysis of an Interferon Gene from the Zebrafish, *Danio rerio*. *J Virol.* 2003 Feb; 77: 1992-2002. doi: 10.1128/jvi.77.3.1992-2002.2003.

116- Rokenes TP, Larsen R, Robertsen B. Atlantic salmon ISG15: expression and conjugation to cellular proteins in response to interferon, double-stranded RNA and virus infections. *Molecular Immunology.* 2007 Feb; 44: 950–959. doi: 10.1016/j.molimm.2006.03.016.

117- Ooi EL, Verjan N, Hirono I, Nochi T, Kondo H, Aoki T, Kiyono H, Yuki Y. Biological characterisation of a recombinant Atlantic salmon type I interferon synthesized in *Escherichia coli*. *Fish and Shellfish Immunology.* 2008 Jun; 24(5): 506–513. doi: 10.1016/j.fsi.2007.10.004.