



Effect of Sodium Nitroprusside (SNP) on Minerals Content of Cabbage *Brassica oleracea var. capitata* L. Grown under Salt Stress

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Abstract: The experiment was conducted during the winter season of 2017-2018 and 2018-2019 at directorates of agriculture in Al-Zubair district, Basrah, Iraq to study the effect of sodium nitroprusside (SNP) at four concentration (0, 50, 100 and 150) μM with number of sprays (once and twice) and three cultivars (Pruktor F1, Luna and Rain ball F1) on mineral content of Nitrogen (N), Phosphorous (P), Potassium (K), Sodium (Na), Potassium/Sodium (K^+/Na^+) ratio, Chloride (Cl^-), Sulfur (S) and Iron (Fe) of leaves. Split-Split Plot design was used with three replicates per treatment. The means of treatments were compared by L.S.D. at 0.05 probability. Results indicated that Pruktor F1 had the highest percentage of N, P, K, K^+/Na^+ ratio for both growing seasons while the highest accumulation of Na and Cl^- in Rain ball F1 cultivar. Plants sprayed twice with SNP had a significant increase in P, K, K^+/Na^+ ratio, S, and Fe comparing with once spray for both growing seasons. PruktorF1 sprayed with SNP at 100 μM twice time was superior in P, K, K^+/Na^+ ratio, S and Fe, whereas, the same cultivar at 150 μM concentration was superior in N for both growing seasons, same cultivar at 100 and 150 μM sprayed twice time gave the lowest percentage of Na^+ and Cl^- , respectively. Also Rain ball F1 at control treatment had the highest accumulation of Na and of Cl^- .

Keywords: Sodium nitroprusside, Salt stress, Nitric oxide, Nitrogen, Phosphorus, Potassium.

Introduction

Cabbage belongs to the Cruciferae family and considers of the favourite winter crop in Iraq and other countries due to its using of salad and cooking, and it is planted in most regions of Iraq and the planted areas were reached 3315 Donums at 2018 with a total production of 6130 tons (Agricultural Statistics Directorate, 2018).

The salinity of irrigation water is one of the most important problems facing agriculture in

arid and semi-arid region of the world and it is one of the main problems that faced the agricultural production in the desert region in Al-Zubair resulting in decreasing growth and photosynthesis efficiency, respiration and availability of nutrients and oxidation as a result of salt stress due to reactive oxygen species (ROS) (Munns & Tester, 2008). Agriculture in Al-Zubair depends on the well waters because of the Lack of surface water, low rainfall and the random use of salty water

leads to negative results on the availability of nutrients through the toxicity of Na⁺ and Cl⁻ ions and the increasing of them with other ions leads to increase the osmotic pressure and imbalance the homeostasis nutrients, therefore methods must be applied in order to increase production and reduce environmental stress (Phocaidés, 2001).

Cabbage is moderately sensitive to salinity and the *salinity has caused negative effects* on production (Ayers, & Wescot, 1985), and it is necessary to use some techniques to reduce the influence of salt stress by using the SNP (donor NO), an inorganic compound (Na₂[Fe(CN)₅NO] .2H₂O) which used to cure from heart disease for human by expanding vascular and it is essential drugs (WHO, 2015). It is one of the important technique which used for regulating most of physiological processes, including the responses to biotic and abiotic stress and increase the tolerance to salt stress by stimulating the antioxidant enzymes, which play a vital role in protection system and increase their tolerance to salt stress resulting in homeostasis nutrients and ionic balance (Nabi *et al.*, 2019; Santisree *et al.*, 2019).

Many studies referred to the spray with SNP compound for plants under salt stress (Molassiotis *et al.*, 2010) and Nitric oxide (NO) increase the activity of plasma membrane H⁺-ATPase in plant exposed to salt stress and led to increase in K⁺/Na⁺ ratio in the tissue of plants to increase adaptation to salt stress (Zhao *et al.*, 2004). NO has a capability to remove *reactive oxygen species*(ROS) and acts as antioxidant by altering the gene expression of antioxidants and thus *protect cells from oxidative damage* caused by salt stress (Arasimowicz & Floryszak-Wieczorek, 2007).

An experiment aimed to study the effect of SNP and the number of sprays on leaf N, P, K, Na, K⁺/Na⁺ ratio, Cl⁻, S and Fe content for three cultivars of cabbage under salt stress in the desert region southern Iraq.

Materials & Methods

A-field experiment

The experiment was carried out during the winter season of 2017-2018 and 2018-2019 in the tomato development project at Al-Zubair, Directorate of Agriculture of Basrah. Random samples of field soil was taken to estimate some of the chemical and physical properties of it (table 1) and table (2) showed some chemical and physical properties of irrigation water (well water). The soil was plowed twice Perpendicular to a depth 30 cm and left for a month, the field arranged in 18 lines 28.8 meters long and 40 cm wide and 40 cm between the lines and every line was divided into four experimental units with a length 7.2 meter. The field soil fertilized with an organic fertilizer at the rate of 13 tone.Donum⁻¹ and decomposed fertilizer NPK (15: 15: 15 + TE) at the rate 0.5 kg for every unit and covered with the soil of the field with at 10 cm layer. The field was supplied with a drip irrigation system and the line covered with black plastic mulching.

The seeds were sown in styropor trays with 209 holes and were sterilized with beltanol 50% SL and filled with German peat moss, at 10/9/2017 and 9/9/2018 for both growing seasons. The seedlings were transplanted to the field after 45 days from sowing. First spray with the SNP after two weeks of transplanting and second spray after two

Table (1): Physical and chemical properties of the field soil.

Properties of the soil		2017-2018 Season	2018-2019 Season	Type of analysis
Electrical conductivity (ds.m ⁻¹)		7.11	7.40	Page <i>et al.</i> (1982)
Soil pH		7.31	7.55	
Soluble ions (mM)	Na ⁺	24.0	30.0	Richards (1954)
	Ca ⁺⁺	17.75	20.00	
	Mg ⁺⁺	15.0	10.5	
	SO ₄ ⁻	19.62	23.14	Page <i>et al.</i> (1982)
	Cl ⁻	67.00	65.00	Jackson (1958)
	HCO ₃ ⁻	2.6	2.8	Richards (1954)
Available Nitrogen		154	170	
Available phosphorus	mg.Kg ⁻¹	69.02	75.40	Page <i>et al.</i> (1982)
Available potassium		201.0	185.4	
Organic matter (g.Kg ⁻¹)		4.64	5.08	
Soil structure				
Sand		83.0	83.0	Black (1965)
Silt		3.6	3.6	
Clay		13.4	13.4	
Soil texture		Loamy sand texture		

Table (2): Physical and chemical properties of irrigation water (well water).

Properties of the water	Unit	2017-2018 Season	2018-2019 Season
pH	-	7.11	7.35
Electrical conductivity (EC)	ds.m ⁻¹	11.68	16.55
Ca ⁺⁺	mg.L ⁻¹	570.0	480.0
Mg ⁺⁺		409.9	-
Na ⁺		870.9	1009.0
K ⁺		40.5	55.2
Cl ⁻		177.2	211.0
NO ₃ ⁻		8.0	9.0
SO ₄ ⁻		795.0	850.0

weeks from the first spray for Pruktor F1, Luna and Rain ball F1 with concentration 0, 50, 100 and 150 µM.

The treatments were assigned to a completely randomized block design in a split-split plot arrangement with the cultivar in the main plots and spray time in the sub-plots and foliar sprays of SNP in sub-sub plots with three replicates.

The means of treatments were compared by L.S.D. at 0.05 probability.

B-Determination of leaves content of nutrient elements:

1-Total nitrogen (N %) was determined by micro-kjeldal method according to Page *et al.* (1982).

2-Phosphorus (P %) was determined according to the method described by Jackson (1985).

3-Potassium and Sodium (K %, Na %) were determined according to the method described by Page *et al.* (1982).

4- K^+/Na^+ ratio by dividing K^+ % on Na^+ %.

5-Chloride (Cl^- %) was determined according to the method described by Furman (1962).

6-Sulfur (S%) was determined according to a method described by Novozamsky & Eck (1977).

7-Iron (Fe $mg.kg^{-1}$) was determined by using atomic absorption according to a method described by Haynes (1980).

Results & Discussion

Data presented in tables (3-9) explained that Pruktor F1 cultivar had a significant increase in N, P, K, K^+/Na^+ ratio, S and Fe and a significant decrease in Na^+ and Cl^- compared to other cultivars. Plants sprayed with SNP had a significant increase in the contents of N, P, K, K^+/Na^+ ratio, S and Fe compared with control treatment and significant decrease in Na^+ and Cl^- . Plants which sprayed twice with SNP at 100 μM had the best result in P, K, K^+/Na^+ ratio and Fe. Plants which sprayed once with SNP at 150 μM had significant increases in N comparing with control treatment.

The Pruktor F1 cultivar plants that sprayed twice with SNP at 100 μM had highest values of P, K, K^+/Na^+ ratio S and Fe and lowest values of Cl^- . the same cultivar which sprayed once with 150 μM SNP had gave highest N and lowest values of Na, while untreated Rain ball F1 cultivar gave less N, P, K, K^+/Na^+ ratio, S and Fe and highest values of Na and Cl^- for both growing seasons, respectively.

The results presented in table Tables (3, 4, 5, 7, 9 & 10) revealed that Pruktor F1 cultivar was superior compared to other cultivars. Genetic factors of cultivars and its ability to prevent sodium ion uptake to plant roots. Results exhibited accumulation of nutrient elements except for Na^+ and Cl^- when the SNP used well water with high salt (11.68, 16.55 $ds.m^{-1}$ for both growing seasons (table, 2). Spraying SNP twice had a significant effect (tables 4-5 & 7) than once spray (tables 3, 6 & 8) because of the high concentration had unaffected role, so it exhibited the growth (Hayat *et al.*, 2014) and due to joint interaction with ROS, which causes damage and breakdown of cells in more than one location, including the cellular membrane which causes oxidative stress (Belgini & Lamattina, 1999).

Tables (3-5, 7 & 9) revealed a significant decrease in the content of N, P, K, K^+/Na^+ ratio, S and Fe when irrigated with salty water compared with the an increase of N^+ and Cl^- ions. This lack of nitrogen content (table 3) caused a decrease in protein because of decreasing the activity of Nitrate reductase enzyme which affected of the synthesis of protein and total nitrogen (Lopez-Cantarero *et al.*, 1997; Jabeen & Ahmad, 2011), and the lack of water stimulates protease enzyme (Reddy & Vora, 1985) and the competition between chloride and nitrate ion the exhibited nitrate and transporter because of toxic effect of salt ions (Lin *et al.*, 1997) that led to accumulation of Cl^- in leaves (Dean-Drummond, 1986) and it may also be due to the change in permeability properties of the plasma membrane, which

affected by membrane proteins (Meloni *et al.*, 2004).

The decrease of P in untreated plants (Table 4) were due to the competition between Cl^- and H_2PO_4^- ions that Cl^- ion acts on decrease absorption H_2PO_4^- by the plant (Pessarkli, 1999) and the reducing of phosphorous translocation from root to vegetative growth (Martinez & Luchli, 1994). The salinity leads to reduction of root growth and its movement in soil and because of limiting movement of P lead s to reduce its adsorption (Al-Taey *et al.*, 2017). The reduction of K (Table 5) in untreated plants were due to the competition between Na^+ and K^+ ion on the absorption sites in roots and transporter proteins which transport Na^+ lasted of K^+ (Ashley *et al.*, 2006).

Data presented in table (6) revealed the reduction in K^+ in untreated plants because of well water that lead to reduce K^+ absorption because of its effect on cellular organelles and transporter of Na^+ and K^+ and H^+ pumps which generate the transporter forces ion in the cells (Zhu, 2003) and the reduction in water absorption was due to the high osmotic pressure in zone root that lead to reduced absorption on K^+ ion (Cuartero & Fernande-Munoz, 1999) and that leads to reduction in the K^+/Na^+ ratio (Table 7).

The high percentage of Na^+ and Cl^- in untreated control (Table 6 & 8) because of their high concentration in a growth medium that lead to an increase of their absorption in plants.

Data presented in table (9) explained a significant decrease in S in untreated plants

were due to the high salt in soil that lead to alkaline pH and saturated with calcium ion (Dougrameji & Al-Rawi, 1972) and that effect on the availability of nutrients which important for plant growth. There was a significant decrease in the Iron element of untreated plants (Table, 10) because the irrigation water salinity lead to change the pH in soil and Fe sensitive to pH and that lead to reduce its availability for plant and then reduce its level in leaves and there was another reason of the toxic effect of Na^+ and Cl^- on plasma membrane of root cells, which lead to reduce its ability for absorption and finally on absorption of nutrient elements like Fe (Passarakli, 1999).

Tables (3-5 & 7) showed a significant increase in the percentage of nutrient elements which treated with the SNP because NO plays a vital role in alleviating of high salt in plant tissue and reduce the nutrient absorption. SNP leads to increase gene expression to H^+ -ATPase in plasma membrane and that lead to raising K^+/Na^+ in cells cytoplasm (Zhao *et al.*, 2004) and showed the addition of SNP on *Zea mays* caused increasing the activity of H^+ -ATPase in tonoplast and Na^+/H^+ transporter and enable Na^+ for passing and SNP improve macronutrient elements contents like Fe (Graziano *et al.*, 2002).

NO plays a physiological role in improving the transport of Fe from root to vegetative system, also, it made many nutrient elements can be absorbed by Iron-regulated transporter 1 (IRT1), which lead to an increase gene expression for them by NO (Connolly *et al.*, 2002).

Table (3): Effect of cultivars, sprays and concentration of SNP on N-content (%) in leaves.

SNP μM	Sprays	2017 – 2018 Season				2018 – 2019 Season			
		Cultivars			SNP × Sprays	Cultivars			SNP × Sprays
		Prukt or F1	Luna	Rain ball F1		Prukt or F1	Luna	Rain ball F1	
0	Once time	2.96	2.77	2.44	2.73	2.91	2.75	2.38	2.61
	Twice time	2.99	2.81	2.49	2.76	2.97	2.79	2.43	2.64
50	Once time	3.27	3.14	2.99	3.13	3.45	3.11	2.91	3.03
	Twice time	3.84	3.38	3.35	3.52	3.43	3.34	3.27	3.44
100	Once time	3.64	3.35	3.24	3.41	3.77	3.30	3.25	3.35
	Twice time	3.51	3.19	3.14	3.28	3.51	3.16	3.11	3.23
150	Once time	4.08	3.69	3.59	3.78	4.03	3.65	3.49	3.67
	Twice time	3.46	3.15	3.14	3.25	3.45	3.16	3.08	3.18
LSD 0.05		0.09		0.05		0.09		0.06	
Effect of cultivars		3.47	3.19	3.05	Effect of SNP	3.34	3.10	2.99	Effect of SNP
LSD 0.05		0.02		0.03					
Cultiva rs × SNP	0	2.98	2.79	2.46	2.74	2.81	2.67	2.38	2.62
	50	3.55	3.26	3.17	3.33	3.44	3.17	3.09	3.23
	100	3.57	3.27	3.19	3.35	3.47	3.22	3.18	3.29
	150	3.77	3.42	3.37	3.52	3.64	3.34	3.30	3.43
LSD 0.05		0.06		0.04		0.05		0.03	
				Effect of Sprays				Effect of Sprays	
Cultiva rs × Sprays	Once time	3.47	3.24	3.06	3.26	3.36	3.14	3.00	3.17
	Twice time	3.45	3.13	3.03	3.21	3.33	3.06	2.97	3.12
LSD 0.05		0.03		0.02		*NS		NS	

*NS: *not significant*

Table (4): Effect of cultivars, sprays and concentration of SNP on P-content (%) in leaves.

SNP μM	Sprays	2017 – 2018 Season				2018 – 2019 Season			
		Cultivars			SNP × Sprays	Cultivars			SNP × Sprays
		Prukt or F1	Luna	Rain ball F1		Prukto r F1	Luna	Rain ball F1	
0	Once time	0.330	0.302	0.280	0.304	0.314	0.284	0.269	0.289
	Twice time	0.329	0.306	0.284	0.306	0.310	0.293	0.275	0.293
50	Once time	0.383	0.356	0.333	0.358	0.364	0.341	0.322	0.342
	Twice time	0.466	0.434	0.411	0.437	0.452	0.417	0.400	0.423
100	Once time	0.441	0.413	0.393	0.415	0.423	0.398	0.380	0.401
	Twice time	0.564	0.525	0.496	0.528	0.540	0.507	0.487	0.512
150	Once time	0.504	0.456	0.451	0.470	0.481	0.438	0.438	0.452
	Twice time	0.459	0.479	0.440	0.459	0.444	0.462	0.439	0.448
LSD 0.05		0.012		0.007		0.015		0.008	
Effect of cultivars		0.434	0.409	0.386	Effect of SNP	0.416	0.392	0.376	Effect of SNP
LSD 0.05		0.006		0.009					
Cultiva rs × SNP	0	0.329	0.304	0.282	0.305	0.312	0.288	0.272	0.291
	50	0.425	0.395	0.372	0.397	0.408	0.379	0.361	0.383
	100	0.503	0.469	0.444	0.472	0.482	0.453	0.434	0.456
	150	0.481	0.468	0.445	0.465	0.463	0.450	0.438	0.450
LSD 0.05		0.009		0.005		0.011		0.006	
				Effect of Sprays				Effect of Sprays	
Cultiva rs × Sprays	Once time	0.414	0.382	0.364	0.387	0.395	0.365	0.352	0.371
	Twice time	0.455	0.436	0.408	0.433	0.437	0.420	0.400	0.419
LSD 0.05		0.006		0.003		NS		0.004	

Table (5): Effect of cultivars, sprays and concentration of SNP on K content (%) in leaves.

SNP μM	Sprays	2017 – 2018 Season				2018 – 2019 Season			
		Cultivars			SNP × Sprays	Cultivars			SNP × Sprays
		Prukt or F1	Luna	Rain ball F1		Prukt or F1	Luna	Rain ball F1	
0	Once time	1.92	1.76	1.62	1.77	1.79	1.60	1.53	1.64
	Twice time	1.90	1.73	1.64	1.76	1.78	1.62	1.54	1.65
50	Once time	2.42	2.31	1.99	2.24	2.19	1.87	1.78	1.95
	Twice time	3.03	2.82	2.44	2.76	2.83	2.62	2.28	2.58
100	Once time	2.88	2.61	2.38	2.62	2.68	2.42	2.29	2.46
	Twice time	3.77	3.15	2.82	3.25	3.52	3.05	2.61	3.06
150	Once time	3.61	2.90	2.64	3.05	3.30	2.85	2.46	2.87
	Twice time	2.55	2.56	2.28	2.46	2.38	2.30	2.21	2.29
LSD 0.05		0.09		0.05		0.22		0.12	
Effect of cultivars		2.76	2.48	2.23	Effect of SNP	2.56	2.29	2.09	Effect of SNP
LSD 0.05		0.04		0.09		0.09		0.09	
Cultiva rs × SNP	0	1.91	1.75	1.63	1.76	1.78	1.61	1.54	1.64
	50	2.72	2.57	2.21	2.50	2.51	2.24	2.03	2.26
	100	3.32	2.88	2.60	2.93	3.10	2.74	2.45	2.76
	150	3.08	2.72	2.47	2.76	2.84	2.57	2.34	2.58
LSD 0.05		0.07		0.04		NS		0.09	
				Effect of Sprays		C		Effect of Sprays	
Cultiva rs × Sprays	Once time	2.71	2.40	2.16	2.42	2.49	2.18	2.02	2.23
	Twice time	2.81	2.57	2.30	2.56	2.63	2.40	2.16	2.40
LSD 0.05		NS		0.03		NS		0.06	

Table (6): Effect of cultivars, sprays and concentration of SNP on Na content (%) in leaves.

SNP μM	Sprays	2017 – 2018 Season				2018 – 2019 Season			
		Cultivars			SNP × Sprays	Cultivars			SNP × Sprays
		Prukt or F1	Luna	Rain ball F1		Prukt or F1	Luna	Rain ball F1	
0	Once time	1.693	1.835	2.035	1.854	1.879	1.990	2.113	1.994
	Twice time	1.690	1.852	2.024	1.855	1.873	1.962	2.144	1.993
50	Once time	1.560	1.711	1.898	1.723	1.706	1.865	1.968	1.846
	Twice time	1.207	1.424	1.631	1.421	1.339	1.634	1.816	1.597
100	Once time	1.309	1.572	1.702	1.528	1.572	1.735	1.886	1.731
	Twice time	0.986	1.280	1.439	1.235	1.215	1.396	1.509	1.373
150	Once time	1.117	1.374	1.575	1.355	1.394	1.563	1.725	1.561
	Twice time	0.871	1.079	1.310	1.087	1.088	1.201	1.409	1.233
LSD 0.05		NS			0.039	0.052			0.031
Effect of cultivars		1.304	1.516	1.702	Effect of SNP	1.508	1.668	1.821	Effect of SNP
LSD 0.05		0.023				0.024			
Cultiva rs × SNP	0	1.691	1.843	2.029	1.855	1.876	1.976	2.128	1.993
	50	1.383	1.568	1.765	1.572	1.523	1.749	1.892	1.721
	100	1.148	1.426	1.570	1.381	1.393	1.566	1.697	1.552
	150	0.994	1.227	1.442	1.221	1.241	1.382	1.567	1.397
LSD 0.05		0.044			0.027	0.036			0.021
					Effect of Sprays				Effect of Sprays
Cultiva rs × Sprays	Once time	1.420	1.623	1.802	1.615	1.638	1.788	1.923	1.783
	Twice time	1.188	1.409	1.601	1.399	1.379	1.549	1.719	1.3549
LSD 0.05		NS			0.024	NS			0.020

Table (7): Effect of cultivars, sprays and concentration of SNP on K⁺/Na⁺ ratio in leaves.

SNP μM	Sprays	2017 – 2018 Season				2018 – 2019 Season			
		Cultivars			SNP × Sprays	Cultivars			SNP × Sprays
		Prukt or F1	Luna	Rain ball F1		Prukt or F1	Luna	Rain ball F1	
0	Once time	1.135	0.961	0.795	0.964	0.951	0.805	0.725	0.827
	Twice time	1.123	0.937	0.808	0.956	0.952	0.826	0.719	0.832
50	Once time	1.549	1.351	1.048	1.316	1.285	1.002	0.906	1.064
	Twice time	2.512	1.983	1.497	1.997	2.112	1.603	1.254	1.656
100	Once time	2.200	1.658	1.399	1.752	1.708	1.394	1.214	1.439
	Twice time	3.832	2.464	1.962	2.753	2.900	2.189	1.728	2.272
150	Once time	3.233	2.111	1.679	2.341	2.369	1.820	1.427	1.872
	Twice time	2.931	2.372	1.744	2.349	2.184	1.912	1.569	1.888
LSD 0.05		0.150		0.090		0.129		0.076	
Effect of cultivars		2.314	1.729	1.366	Effect	1.808	1.444	1.193	Effect
LSD 0.05		0.071		of SNP		0.053		of SNP	
Cultiva rs × SNP	0	1.129	0.949	0.802	0.960	0.952	0.815	0.722	0.829
	50	2.030	1.667	1.273	1.656	1.698	1.302	1.080	1.360
	100	3.016	2.061	1.681	2.253	2.304	1.791	1.471	1.855
	150	3.082	2.242	1.711	2.345	2.277	1.866	1.498	1.880
LSD 0.05		0.104		0.060		0.091		0.055	
					Effect of Sprays				Effect of sprays
Cultiva rs × Sprays	Once time	2.029	1.520	1.230	1.593	1.578	1.255	1.068	1.300
	Twice time	2.600	1.939	1.503	2.014	2.037	1.632	1.318	1.662
LSD 0.05		0.090		0.062		0.064		0.042	

Table (8): Effect of cultivars, sprays and concentration of SNP on Cl content (%) in leaves.

SNP μM	Sprays	2017 – 2018 Season				2018 – 2019 Season			
		Cultivars			SNP × Sprays	Cultivars			SNP × Sprays
		Prukt or F1	Luna	Rain ball F1		Prukt or F1	Luna	Rain ball F1	
0	Once time	4.53	4.69	4.79	4.67	4.83	4.97	5.08	4.96
	Twice time	4.42	4.56	4.50	4.49	4.69	4.83	4.77	4.76
50	Once time	4.07	4.24	4.27	4.20	4.33	4.49	4.54	4.45
	Twice time	3.53	3.93	3.77	3.75	3.75	4.16	3.99	3.97
100	Once time	3.60	3.78	3.72	3.70	3.83	3.99	3.95	3.92
	Twice time	2.83	3.30	3.26	3.13	2.99	3.48	3.45	3.31
150	Once time	3.23	3.43	3.49	3.38	3.43	3.62	3.67	3.57
	Twice time	2.89	3.32	3.21	3.14	3.04	3.52	3.39	3.32
LSD 0.05		0.13		0.08		0.13		0.08	
Effect of cultivars		3.64	3.91	3.88	Effect of SNP	3.86	4.13	4.10	Effect of SNP
LSD 0.05		0.06		0.07					
Cultiva rs × SNP	0	4.47	4.63	4.65	4.58	4.76	4.90	4.93	4.86
	50	3.80	4.09	4.02	3.97	4.04	4.33	4.26	4.21
	100	3.21	3.54	3.49	3.41	3.41	3.73	3.70	3.61
	150	3.06	3.38	3.35	3.26	3.23	3.57	3.53	3.45
LSD 0.05		NS		0.05		0.10		0.05	
				Effect of Sprays				Effect of sprays	
Cultiva rs × Sprays	Once time	3.86	4.04	4.07	3.99	4.10	4.27	4.31	4.23
	Twice time	3.42	3.78	3.69	3.63	3.62	4.00	3.90	3.84
LSD 0.05		NS		0.06		NS		0.06	

Table (9): Effect of cultivars, sprays and concentration of SNP on S content (%) in leaves.

SNP μM	Sprays	2017 – 2018 Season				2018 – 2019 Season			
		Cultivars			SNP × Sprays	Cultivars			SNP × Sprays
		Prukt or F1	Luna	Rain ball F1		Prukt or F1	Luna	Rain ball F1	
0	Once time	0.417	0.390	0.375	0.394	0.404	0.377	0.367	0.383
	Twice time	0.397	0.391	0.370	0.386	0.387	0.382	0.360	0.376
50	Once time	0.497	0.460	0.450	0.469	0.486	0.448	0.429	0.454
	Twice time	0.561	0.551	0.541	0.551	0.542	0.540	0.529	0.537
100	Once time	0.551	0.542	0.525	0.539	0.533	0.530	0.511	0.525
	Twice time	0.731	0.681	0.620	0.677	0.717	0.670	0.604	0.664
150	Once time	0.626	0.639	0.582	0.617	0.611	0.632	0.566	0.600
	Twice time	0.675	0.648	0.601	0.641	0.659	0.639	0.586	0.628
LSD 0.05		0.028		0.016		0.020		0.012	
Effect of cultivars		0.557	0.538	0.508	Effect of SNP	0.542	0.526	0.494	Effect of SNP
LSD 0.05		0.013		0.008		0.008		0.008	
Cultiva rs × SNP	0	0.407	0.390	0.373	0.390	0.396	0.379	0.364	0.380
	50	0.529	0.506	0.495	0.510	0.514	0.494	0.479	0.496
	100	0.641	0.612	0.573	0.608	0.625	0.600	0.558	0.594
	150	0.651	0.643	0.592	0.629	0.635	0.631	0.576	0.614
LSD 0.05		0.020		0.012		0.014		0.009	
				Effect of Sprays				Effect of Sprays	
Cultiva rs × Sprays	Once time	0.523	0.508	0.482	0.504	0.509	0.495	0.468	0.491
	Twice time	0.591	0.568	0.533	0.564	0.576	0.558	0.520	0.551
LSD 0.05		NS		0.008		NS		0.007	

Table (10): Effect of cultivars, sprays and concentration of SNP on Fe content (mg.Kg⁻¹ D W) in leaves.

SNP μM	Sprays	2017 – 2018 Season				2018 – 2019 Season			
		Cultivars			SNP × Sprays	Cultivars			SNP × Sprays
		Pruktor F1	Luna	Rain ball F1		Prukt or F1	Luna	Rain ball F1	
0	Once time	45.32	40.78	45.32	40.93	42.77	38.00	34.71	38.49
	Twice time	44.46	40.71	44.46	40.81	41.37	38.90	35.00	38.42
50	Once time	51.36	45.58	51.36	46.57	47.97	42.77	40.21	43.65
	Twice time	66.18	59.59	66.18	60.61	62.07	51.23	51.97	55.09
100	Once time	60.15	57.44	60.15	57.26	55.60	48.02	47.77	50.47
	Twice time	77.03	69.60	77.03	69.95	74.36	61.06	59.45	64.96
150	Once time	69.32	64.72	69.32	64.59	65.11	53.04	53.93	57.36
	Twice time	67.31	60.36	67.31	61.53	62.87	54.00	54.49	57.12
LSD 0.05		1.91		1.14		1.44		0.97	
Effect of cultivars		60.14	54.85	50.86	Effect	56.52	48.38	47.19	Effect
LSD 0.05		0.64		of SNP		0.58		of SNP	
Cultiva rs × SNP	0	44.89	40.74	36.99	40.87	42.07	38.45	34.85	38.46
	50	58.77	52.58	49.43	53.59	55.02	47.00	46.09	49.37
	100	68.59	63.52	58.69	63.60	64.98	54.54	53.61	57.71
	150	68.31	62.54	58.34	63.06	63.99	53.52	54.21	57.24
LSD 0.05		NS		0.82		0.86		0.50	
				Effect of Sprays				Effect of Sprays	
Cultiva rs × Sprays	Once time	56.53	52.13	48.36	52.34	52.86	45.46	44.16	47.49
	Twice time	63.74	57.56	53.37	58.23	60.17	51.30	50.23	53.90
LSD 0.05		0.89		0.65		NS		0.84	

Conclusions

The results showed that sodium nitroprusside (SNP) at 100 μM Sprays twice-time enhanced the tolerance of cabbage cv. Pruktor F1 to salt stress by

improving accumulation of P, K, Fe and S and increase K⁺/Na⁺ ratio in leaves.

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References

- Agricultural Statistics Directorate (2018). Central Statistical Organization. Min. Planning, Iraq.
- Al-Taey, D.K.A.; Al-Janabi, A.H. & Rachid, A.M. (2017). Effect of water salinity and organic & mineral fertilizers on the growth and some contents of leave nutrients of cabbage (*Brassica oleracea* var. *capitata* L.). J. Univ. Babylon Pure Appl. Sci., 25(6): 2046- 2064.
- Arasimowicz, M. & Floryszak-Wieczorek, J. (2007). Nitric oxide as a bioactive signaling molecule in plant stress responses. Plant Sci., 172: 876-887.
- Ashley, M.K.; Grant, M. & Grabov, A. (2006). Plant responses to potassium deficiencies: a role for potassium transport proteins. J. Exp. Bot., 57(2): 425-436.
- Ayers, R.S. & Wescot, D.V. (1985). Water quality for agriculture. FAO Irrigation and Drainage Paper 29. FAO, Roma, 174pp.
- Belgini, M.V. & Lamattina, L. (1999). Nitric oxide counteracts cytotoxic process mediated by reactive oxygen species in plant tissues. Planta, 208: 337-44.
- Black, C.A. (1965). Method of Soil Analysis. Part (1). Physical properties. Am. Soc. Agron. Inc. Publisher, Madison, Wisconsin: 770pp..
- Connolly, E.; Fett, J.P. & Guerinot, M. L. (2002). Expression of the IRT1 metal transporter is controlled by metals at the levels of transcript and protein accumulation. Plant Cell, 14: 1347-1357.
- Cuartero, J. & Fernande-Munoz, R. (1999). Tomato and salinity. Sci. Horticult., 78: 83-125 .
- Dean-Drummond, C.E. (1986). A comparison of regulatory effects of chloride on nitrate uptake, and on chloride uptake into *Pisum sativum* seedlings. Physiol. Plant, 66: 115-126.
- Duogrameji, J. & Al-Rawi, J. (1972). Partical size dispersion method in some Iraqi soil. Zeitschrift für Pflanzenährung und Bodenkunde, 131(1): 38-42.
- Furman, N.H. (1962). Standard Method of Chemical Analysis. 6th ed. D. Van Nostrand, Co., Inc., Princeton, N.J.: 365pp.
- Graziano, M.; Beligni, M.V.L. & Lamattina, L. (2002). Nitric oxide improves internal iron availability in plants. Plant Physiol., 130: 1852-1859.
- Hayat, S.; Yadav, S.; Alyemeni, M.N. & Ahmad, A. (2014). Effect of sodium nitroprusside on the germination and antioxidant activities of tomato (*Lycopersicon esculentum* Mill). Bulg. J. Agric. Sci., 20(1): 156-160.
- Haynes, R.J. (1980). Ion exchange properties of roots and ionic interactions within the root apoplasm: Their role in

- ion accumulation by plants. Bot. Rev., 46(1): 75-99.
- Jabeen, N. & Ahmad, R. (2011). Foliar application of potassium nitrate affects the growth and nitrate reductase activity in sunflower and safflower leaves under salinity. Not. Bot. Hort. Agrob., 39 (2): 172-178.
- Jackson, M.L. (1958). Soil chemical analysis. Prentice Hall. Inc. Englewood Cliffs, N. J.: 598pp.
- Lin, H.; Sandra, S.S. & Schumaker, K.S. (1997). Salt sensitivity and the activities of the H-ATPase in cotton seedlings. Crop Sci., 37: 190-197.
- Lopez-Cantarero, I.; Ruiz, J.M.; Hernandez, J. & Romero, L. (1997). Nitrogen metabolism and yield response to increase in nitrogen-phosphorus fertilization; Improvement in greenhouse cultivation of eggplant (*Solanum melongena*). J. Agric. Food Chem., 45: 4227-4231.
- Martinez, V. & Lauchli, A. (1994) . Salt-induced inhibition of phosphate uptake in plants of cotton (*Gossypium hirsutum* L.). New Phytol., 125: 609-614.
- Meloni, D.A.; Gulotta, M.R.; Martinez, C. A. & Oliva, M.A. (2004). The effects of salt stress on growth, nitrate reduction and proline and glycinebetaine accumulation in *Prosopis alba*. Braz. J. Plant Physiol., 16(1): 39-46.
- Molassiotis, A.; Tanou, G. & Diamantidis, G. (2010). NO says more than 'YES' to salt tolerance: salt priming and systemic nitric oxide signaling in plants. Plant Signalling and Behavior, 5: 209-212.
- Munns, R. & Tester, M. (2008). Mechanisms of salinity tolerance. Ann. Rev. Plant Biol., 59: 651-681.
- Nabi, R.B.S.; Tayade, R.; Hussain, A.; Kulkarni, K.; Imran, Q.M.; Mun, B.G. & Yun, B.W. (2019). Nitric oxide regulates plant responses to drought, salinity, and heavy metal stress. Environ. Exp. Bot., 161: 120-133.
- Novozamsky, I. & Eck, R. (1977). Total sulphur determination in plant material. Fresen. J. Anal. Chem., 286 (5): 367-368.
- Page, A.L.; Miller, R.H. & Keeney, D.R. (1982). Method of soil and analysis Part 2, 2nded, Agron. 9. Publisher, Madison, Wisconsin: 1158pp.
- Pessaraki, M. (1999). Handbook of Plant of Crop Stress. 2nd en., Univ. CRC Press, Boca Raton: 1254pp.
- Phocaidis, A. (2001). Handbook on perssured irrigation techniques FAO consultant, Rome, chapter 7, Water quality for irrigation.
- Reddy, M.P. & Vora, A.B. (1985). Effect of salinity on protein metabolism in bajra (*Pennisetum typhoides* S and H) leaves. Indian J. Plant Physiol., 28: 190-195.
- Richards, A. (1954). Diagnosis and improvement of saline and alkali soils. Agriculture Handbook, No. 60, USDA, Washington: 160pp.
- Santisree, P.; Adimulam, S. S.; Sharma, K.; Bhatnagar-Mathur, P. & Sharma, K.

- K. (2019). Insights Into the Nitric Oxide Mediated Stress tolerance in plant. 385-406. In Khan, I.M.R.; Reddy, P.S. Ferrante, A. & Khan, N. (Eds.). *Plant Signaling Molecular: Role and Regulation Under Stressful Environments*. Elsevier: 596pp.
- WHO, World Health Organization. (2015). WHO Model List of Essential Medicines (April 2015). 19th list. www.who.int/selection_medicines/committees/expert/20/EML_2015_Final_amended_AUG2015.pdf?au=1.
- Zhao, L.Q.; Zhang, F. ; Guo, J.K.; Yang, Y. L.; Li, B.B. & Zhang, L. X. (2004). Nitric oxide functions as a signal in salt resistance in the calluses from two ecotypes of reed. *Plant Physiol.*, 134: 849-857.
- Zhu, J.K. (2003). Regulation of ion homeostasis under salt stress. *Curr. Opin. Plant Biol.*, 6: 441-445.